

Decision Support System 2.0: Combining plant resistance with monitoring virulence

IHAR, September 6, 2011

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Outline

- **Phytophthora in perspective**
- DSS 1.0: Umbrella Research Plan Phytophthora: 2003 – 2012
 - Focus on spray management
- Durable resistance: DuRPh research plan: 2006 – 2016
 - Focus on multiple resistance genes
- DSS 2.0: adding monitoring virulence genes



Phytophthora 2002: increasing problem



- Adaptation to cultivar resistance
- New virulence by sexual mating
→ Current R-genes 'broken'
- Increased aggressiveness → rapid epidemics (LP < 3 days).
- Broadening of host plant range
- Legislative problems: → 15 x sprays, environment issue, 50% of all fungicides used in NL



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Co-innovation: Umbrellaplan Phytophthora

Consortium formation in 2003:

- Applied Plant Research
- Plant Research International
- Univ. Dept. Sciences

- Agribusiness (breeders, growers, trade, intermediates)
- Min. Agriculture

Aim: 75% reduction of negative
impact of pesticides in 10 years

Budget: 1.5 M€ per year



Epidemiology → DSS 1.0



■ Research aims:

- factors involved in disease development & spread
- Additional control points in life cycle
- Role of partial resistance of potato

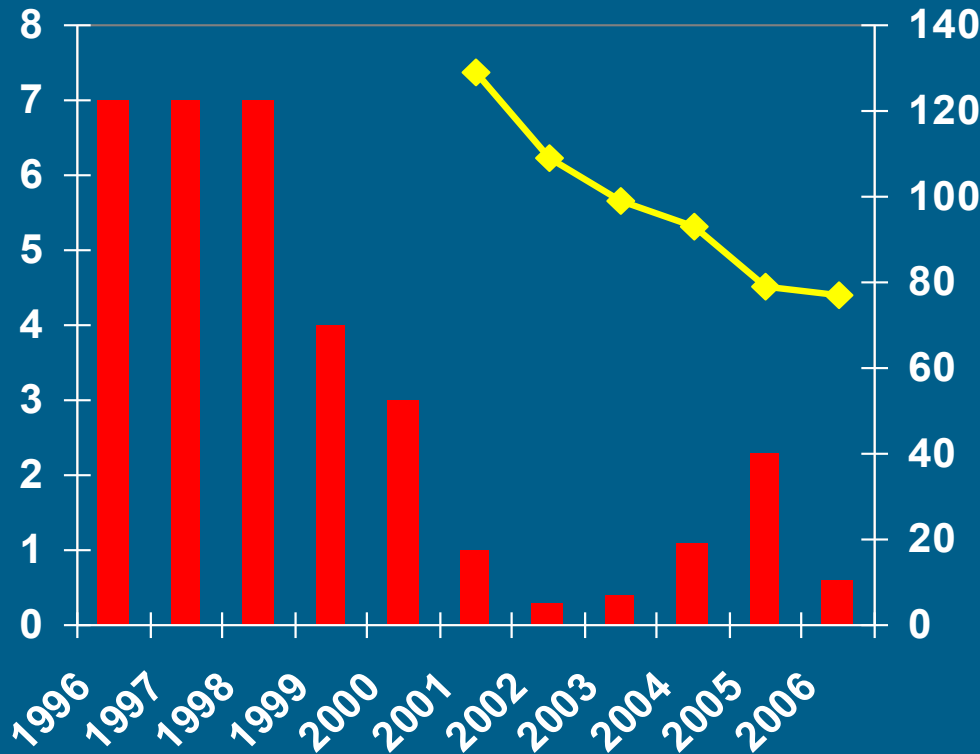
■ Aim for practice:

- What can farmer do for prevention
- Life cycle of disease in practice (what to spray)
- Refining control strategy (when to spray)
- Use of resistance (how much to spray)
- Convincing farmers (communication)

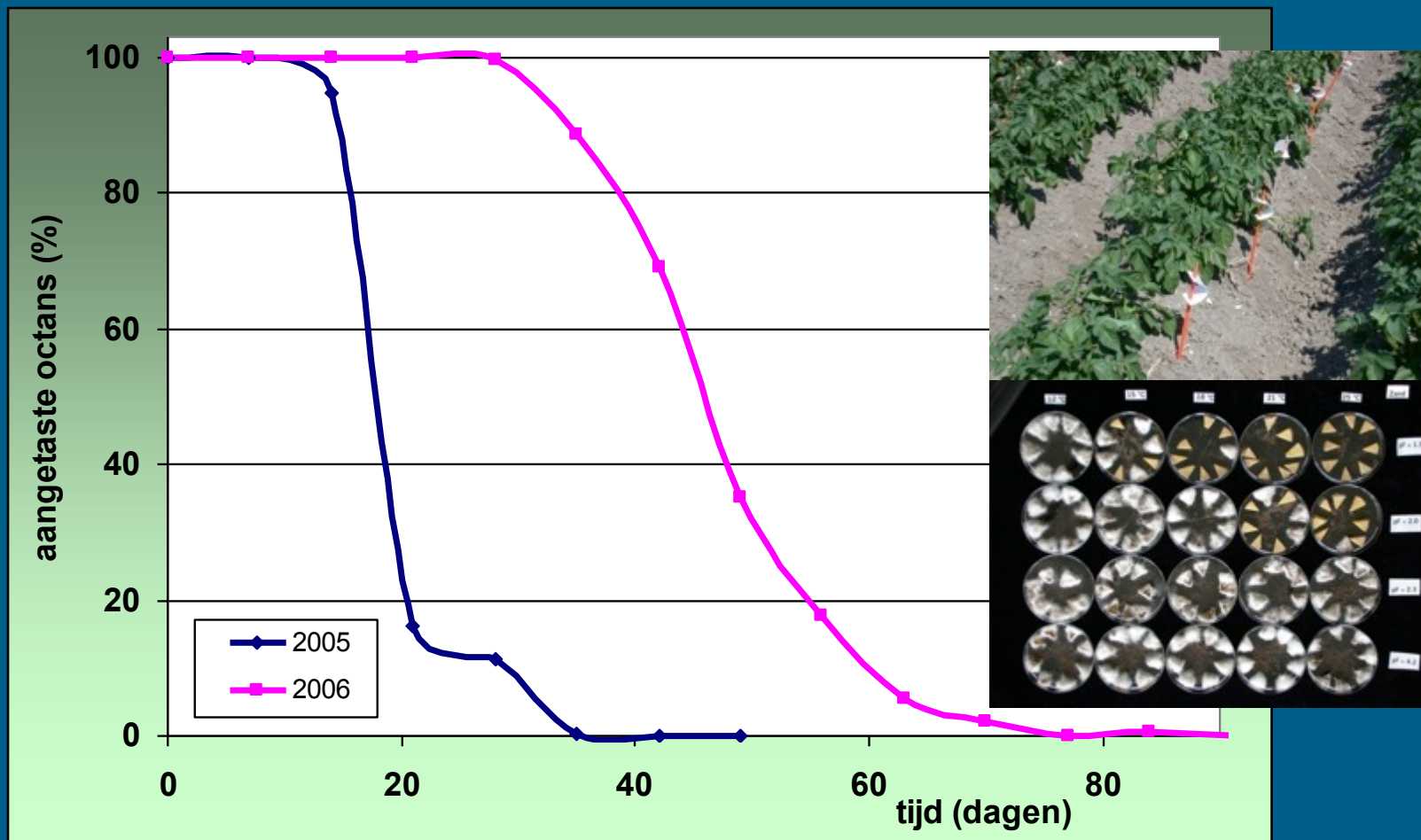
→ Implementation DSS 1.0



Incidence of uncovered dumps: prevention



Survival of sporangia in soil: prevention and what to spray



Control strategy: **What** to spray



cymoxanil

Fubol Gold

Ranman

Ranman

Valbon/Acrobat/Tattoo/Curzate /Sereno/Unikat Pro

Shirlan

Reduced dose rates when possible

Loofgroeifase

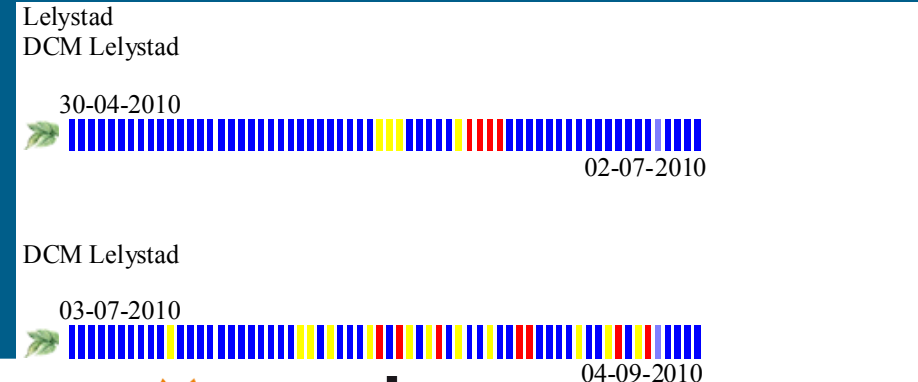
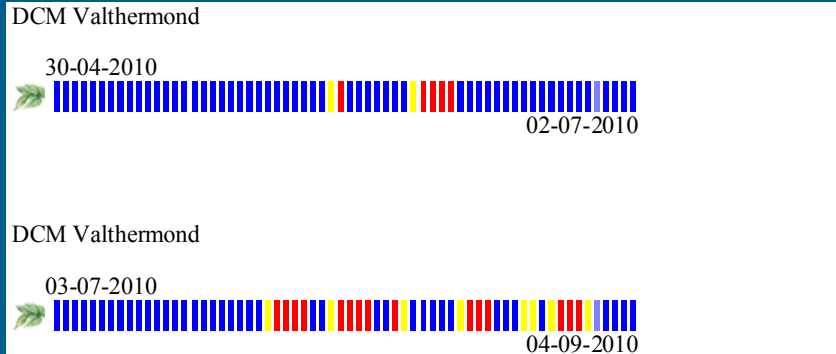
Knolbeschermingsfase



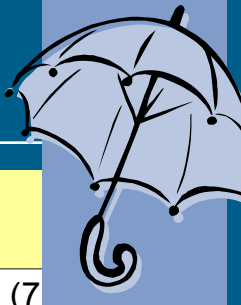
DSS 1.0: When to spray



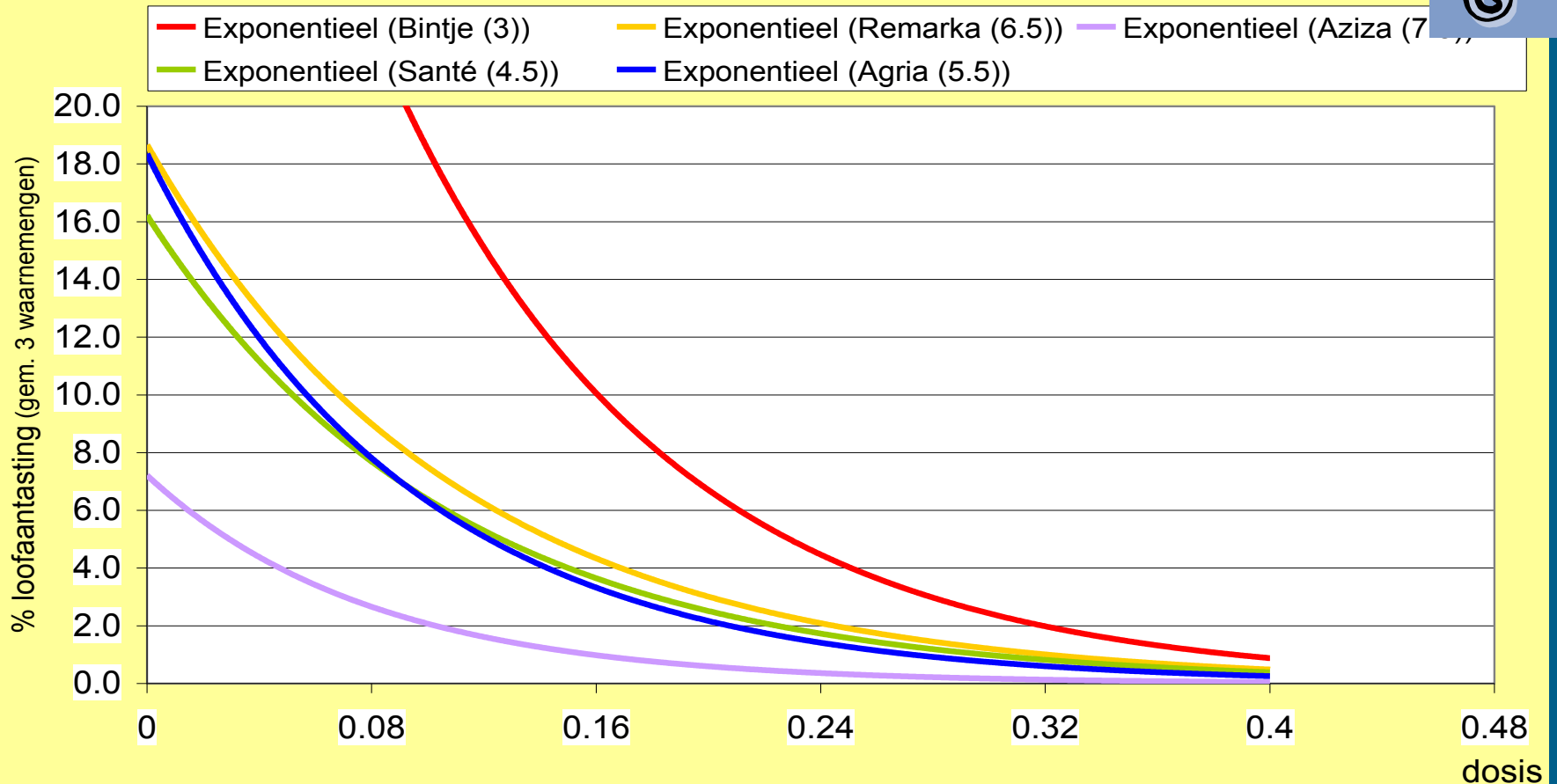
- Accurate weather predictions
- Critical infection circumstances (commercial models)
 - Spores present?
 - Weather conditions favourable for spread and survival of spores?
 - Leaf-wetness favourable for infection?
 - Breakdown time of previously applied fungicide?
 - Canopy development → unprotected areas?



Relative resistance: **how much** to spray



dosis-response grafiek 2004 consumptierassen



Demo trials 2007: communication



- Cultivar resistance & reduced dose rates
- 7 locations
- Spray timing: DSS
- Published weekly
 - Farmers magazines
 - Internet



Results Umbrellaplan Phytophthora



DSS 1.0 implemented:

- Only spray when necessary
- Use the right modern fungicides
- Only spray at optimal conditions
- Use lower dose depending of relative resistance
- Use high tech spray-equipment
- Spray-free zones

More than 75% reduction of negative impact on environment accomplished
BUT: STILL DEPENDANCE!!



New sources for resistance breeding



Results	Genotypes	
Resistant	1209	30%
Intermediate	714	18%
Susceptible	2109	52%
Total resistance data	4032	100%



Durable Resistance against Phytophthora (DuRPh)

DuRPh

duurzame resistentie tegen
Phytophthora in aardappel door
cisgene merkervrije modificatie

Duration 2006 – 2015
Budget €10 M€
Source MinistryLNV



Durable Resistance against Phytophthora

Principles of DuRPh

Cisgene: only use genes from crossable species with *S. tuberosum*

Gene stacking: combining 2-6 genes per cassette,

Maintain varieties: DuRPh maintains present varieties as they are

Deployment: spatial and temporal variation (Flexible varieties)

Marker free: no antibiotics marker

Transparency: communication with all stakeholders concerned

Exploitation: securing intellectual property and sharing breeding rights



PLANT RESEARCH INTERNATIONAL
WAGENINGEN UR



DuRPh

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TRADITIONELE VEREDLING

Crossing



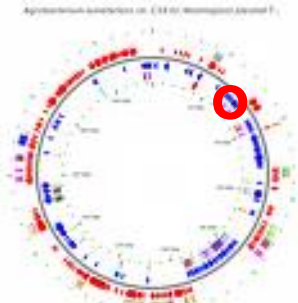
GENTECNOLOGIE



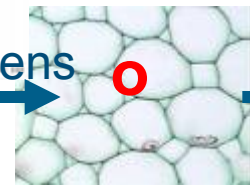
Modification



E. coli



A. tumefaciens



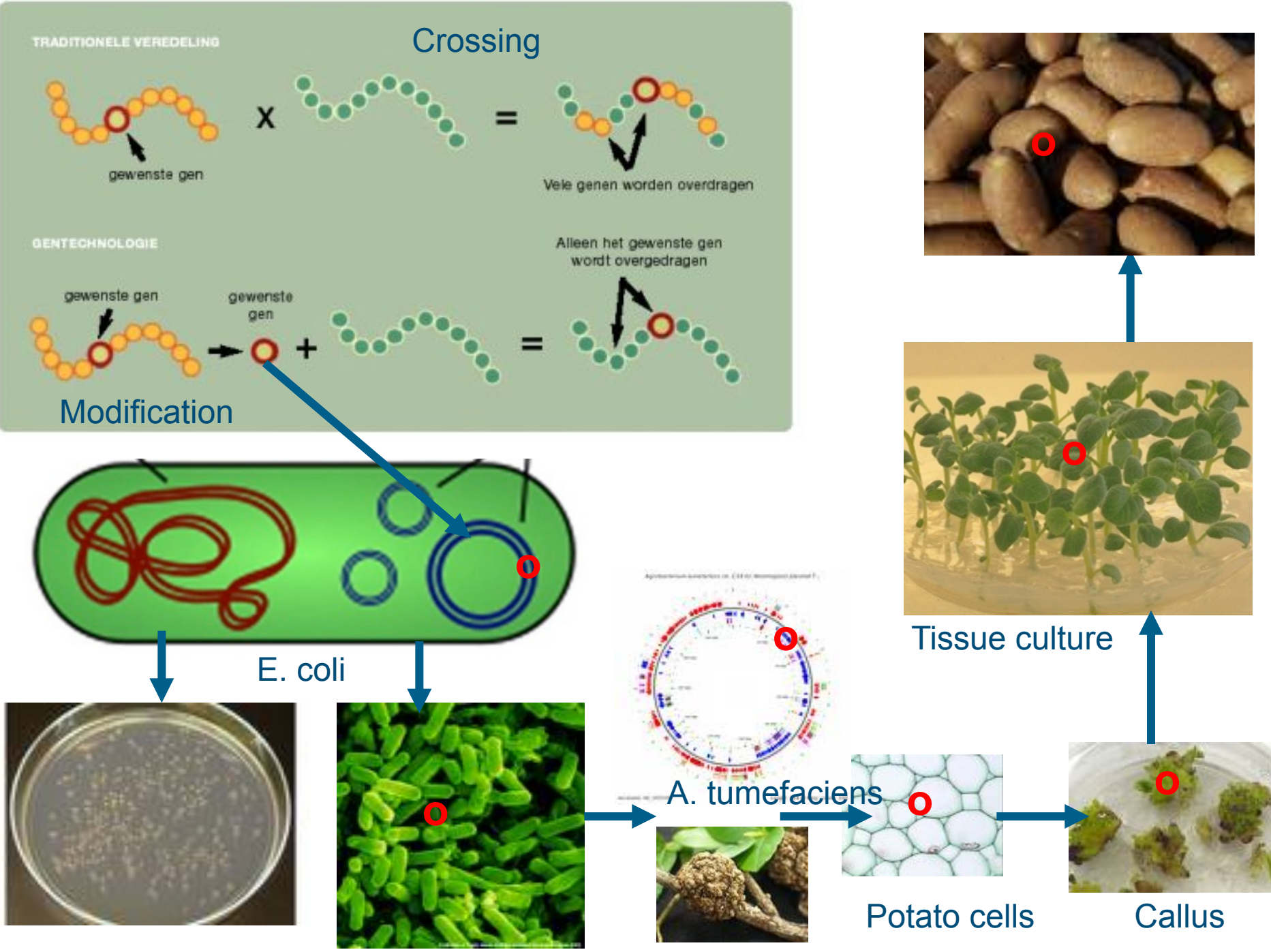
Potato cells



Callus



Tissue culture



Some combinations of R-genes

R-gene cassettes (August 2009)

- Made construct (combination)
 - Rpi-blb1
 - Rpi-blb2
 - R3a
 - Rpi-blb1 + Rpi-blb2
 - Rpi-sto1
 - Rpi-blb3
 - Rpi-blb1 + R3a
 - Rpi-sto1 + R3a
 - Rpi-sto1 + Rpi-blb3

Durable Resistance against Phytophthora



- Selection on
- Resistance level
 - True to type
 - Desirable traits

Genes for potato late blight resistance



Aveka Blb2

Aveka

But...

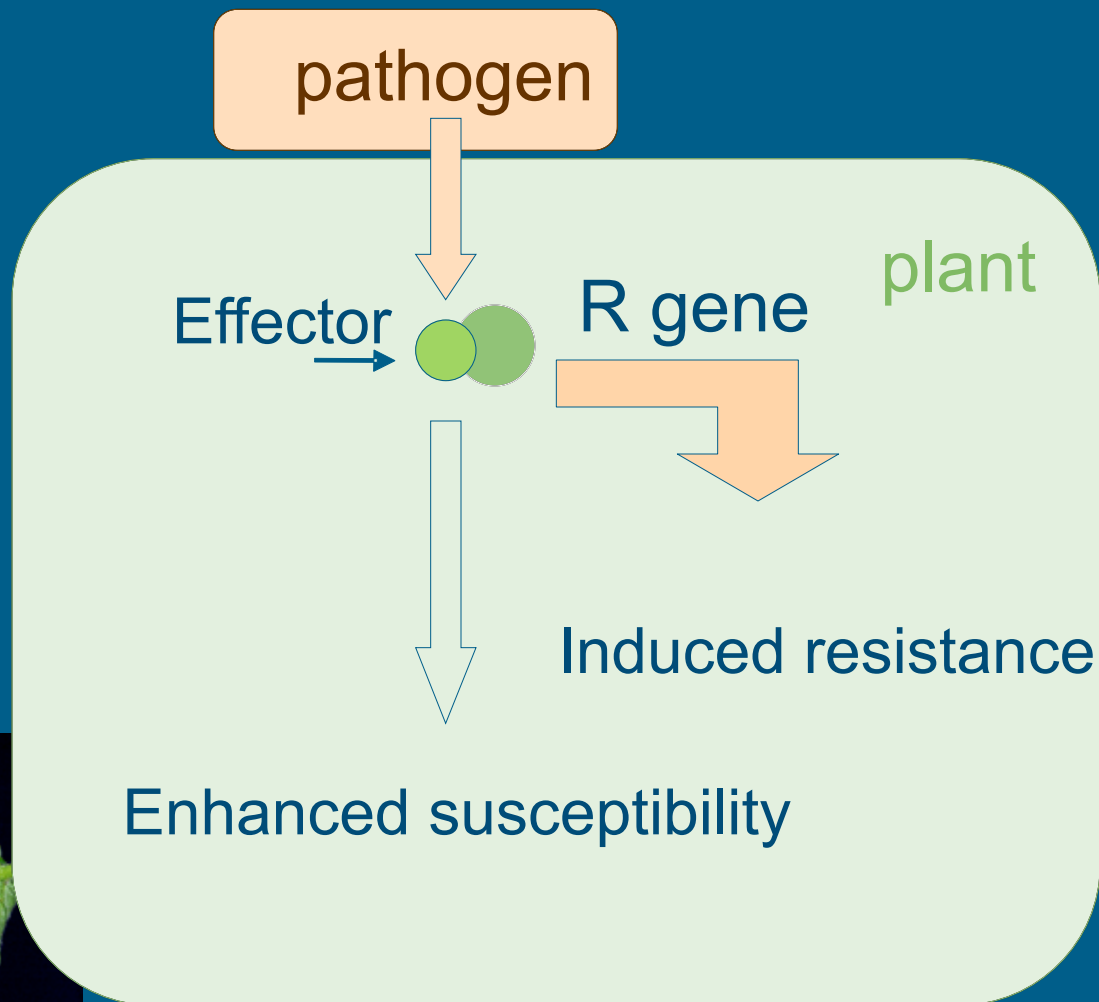
- Will multiple resistance be sustainable?
- When:
 - Temporal and spacial mixing of R-genes
 - Replacing R-genes in time
- Phytophthora will select genotypes that break resistance → precious R-genes will be lost!!

Outline

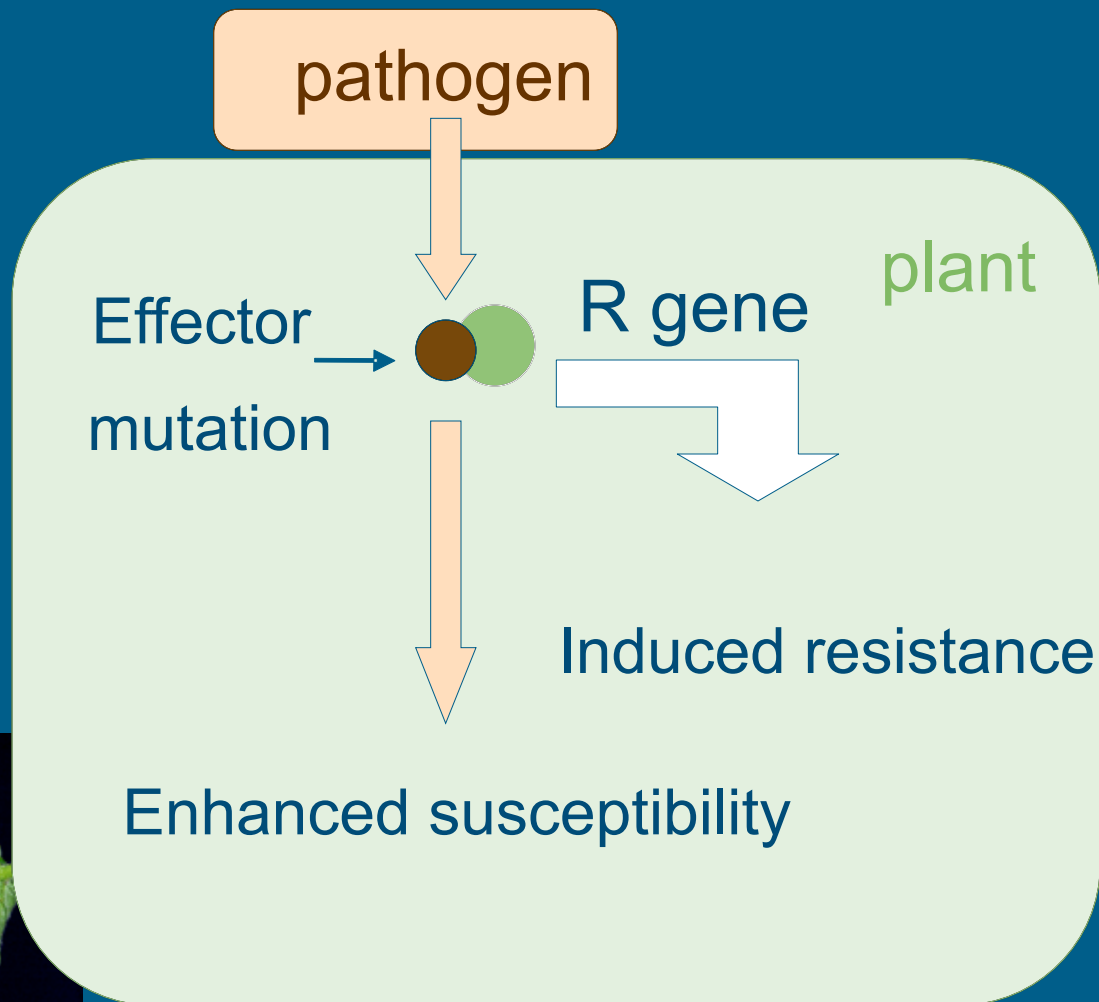
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Effectors and Virulence: a key role for recognition

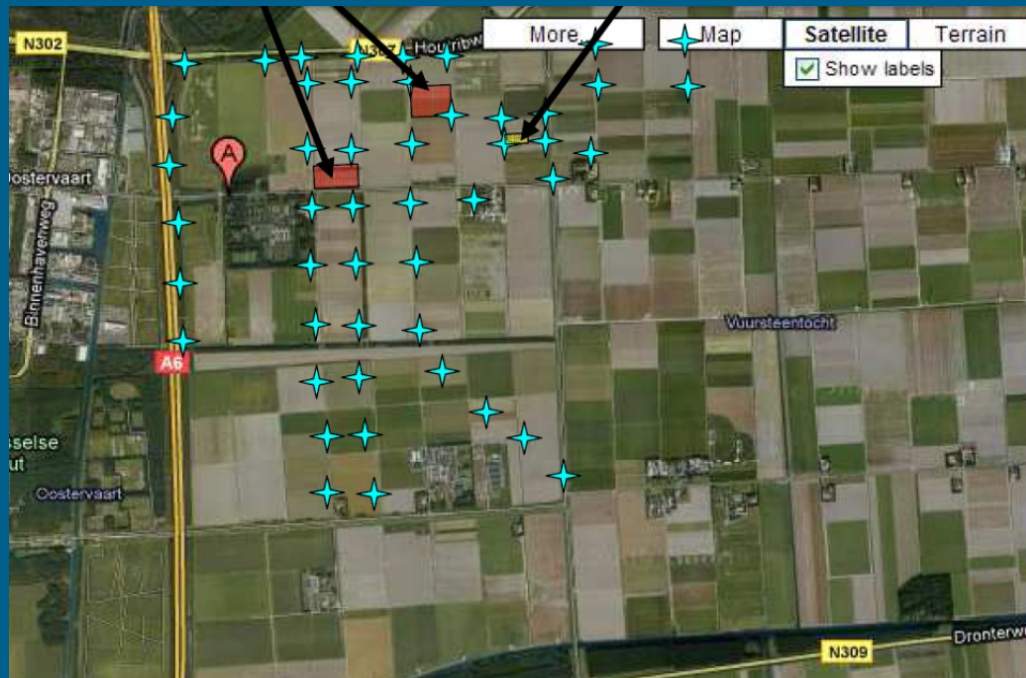


Effectors and Virulence: a key role for recognition



Pilot: Real-time monitoring of Blb1 breakers

- Small bait fields in close to an experimental field
 - Valthermond
 - Lelystad



Pilot: Real-time monitoring of Blb1 breakers



Real-time monitoring of Blb1 breakers

- Advanced TaqMan to monitor alterations in Blb-1 effector gene
- 3 times a week suspicious samples were screened: all NEGATIVE
- But on August 18:
 - 633 suspicious leaf samples were screened **in 8 hr!!!**
 - 75 did not contain *P. infestans*
 - 557 we scored avirulent (intact effector) for Blb1
 - Only ONE sample had a mutated Blb 1 effector gene and is scored virulent

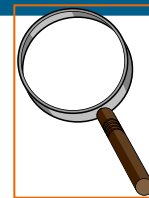
Blb2	LS-17-Bionica		4C10	AVIRULENT
R1R3R10	LS-17-Escort	18-aug-2010	4C11	AVIRULENT
R1R3R10	LS-17-Escort		4C12	AVIRULENT
R1R3R10	LS-18-Escort	18-aug-2010	4D1	AVIRULENT
R1R3R10	LS-18-Escort		4D2	AVIRULENT
Blb2	LS-18-Bionica	18-aug-2010	4D3	VIRULENT
Blb2	LS-18-Bionica		4D4	AVIRULENT
Blb2	LS-19-Bionica		4D5	NO INFESTANS



Principle of DSS 2.0 monitoring in practice

- If a farmer has only Blb 1 cultivars
- Without DSS 1.0: about 15 sprays
- With DSS 1.0: 50% less sprays, with lower dose
- With DSS 2.0 monitoring: result of test in 8 hr, in this year, only ONE SPRAY

→ No breakers, no spray!



- Blb1 is an effective R gene (in the Netherlands)
- Blb1 may be combined with new R genes
- Other effector genes are identified:
 - Avr2, Avr3a, Avr3b, Avr4, AvrVnt1, AvrCh1, AvrBlb1
- Pyramiding could be effective to decrease chance for breaking R
- Monitoring is required to extend the life time of R genes!!!

Towards sustainable control of Phytophthora



Can DSS 2.0 also be developed for other crop – pathosystems??



Thank you for your attention

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